

MicroRNA as non-invasive biomarkers of breast cancer

Saima Sadaf^{*1}, Jawaria Shaheen¹, Kanwal Shabir¹ and M. Waheed Akhtar^{*2}

¹Institute of Biochemistry and Biotechnology, ²School of Biological Sciences, University of the Punjab, Lahore, Pakistan

Abstract: Breast cancer is amongst the most frequently recorded malignancies and the leading cause of cancer deaths in women worldwide including Pakistan. During the recent years, an alarming increase in the incidence and mortality rates of breast cancer in Pakistani women especially in the young girls has been observed. Need for rapid, non-invasive and economical diagnostic procedure therefore is enormous for a population marred with lack of awareness, absence of adequate medical check-up facilities and the socio-economic barriers. Small (18-25 nucleotide long), non-protein-coding microRNAs, which are stably present in the bodily fluid may serve as an ideal class of blood-based biomarkers for cancer detection. This is primarily because the expression of microRNAs is frequently dysregulated in various forms of malignancies including breast cancer. This article reviews the potential of microRNAs to serve as molecular marker of cancer diagnosis, prognosis and/or relapse detection.

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***Authors for Correspondence.** sasadaf@hotmail.com; mwapu@brain.net.pk

INTRODUCTION

Cancer is the leading cause of death and a major public health concern all around the world. Several lines of evidence indicate that the transformation of a normal/healthy cell into malignant tumor and its invasion/metastasis is a multistep process that is accompanied by well-defined genetic, epigenetic and/or molecular abnormalities¹⁻³. The uncontrolled, abnormal growth leads to obstruction of blood vessels, ducts or lymphatics, damages the nerves, causes infections and necrosis of nearby surrounding tissues and eventual death of the patient, if it remains undiagnosed or not countered by treatment.

Whereas cancer cells display some unique characteristics, they are also capable of shedding their nucleic acids into the bloodstream, which may be isolated from serum/plasma and analyzed as surrogate source of tumor DNA. Although healthy subjects do have free circulating DNA in their serum/plasma (15-30 ng/ml) but the mean concentration raises several-folds (upto 180 ng/ml) in patients with different forms of neoplasias³⁻⁴. The mechanism involved in the nucleic acids release is unknown, however, some recent studies have shown interesting results pointing towards the significance of circulating cell-free nucleic acid measurements in early diagnosis of cancers, viz., lung, breast, oral, colon, esophageal and colorectal³⁻⁸.

The potential use of cell-free plasma DNA/RNA for cancer diagnosis is highly attractive as relatively small amount of blood is required for the extraction of nucleic acids and the sample may be drawn

repetitively enabling the analyses of disease progression and treatment response in the patients more objectively. Although El-Hefnawy, *et al.* (2004)⁹ reported that analysis of extracellular plasma RNA, which is an extremely labile molecule, may not be useful for cancer detection or recurrence monitoring but several others remained successful in exploring the usefulness of gene expression profiling of plasma RNA in differentiating the tumor from the healthy conditions⁶⁻⁸.

Amongst the circulating nucleic acids, small (18-25 nucleotide long), non-protein coding RNA i.e., microRNA (miRNA) are exceptionally stable in the RNase rich environment of the bloodstream and other bodily fluids and thus may be well-exploited in blood-based screening of cancer including the breast cancer. This mini-review discusses the potential of microRNAs in breast cancer diagnosis, prognosis and/or relapse detection.

Discovery and functions of microRNA

Small, non-coding *lin-4*, which is involved in the regulation of *lin-14* protein expression, was the first miRNA that was discovered in *Caenorhabditis elegans* during 1993^{10,11}. Few years later, *let-7* (another miRNA), was identified in *C. elegans*¹² and since then several thousand miRNAs have been discovered in different organisms including human beings. The miRBase (www.mirbase.org), a searchable database of miRNAs, contains 28645 published miRNA sequence entries, as per its Release 21 (June 2014) and this

number is increasing with great speed. Interestingly, the studies have shown striking similarities in lin-4 and let-7 from diverse organisms i.e., flies to human beings. This further reflects evolutionary conservation in miRNAs function, which mainly is related to the regulation/control of gene expression in different organisms.

MicroRNAs, like the messenger RNA of higher eukaryotes, are transcribed as long primary-transcripts (pri-miRNA) by type II and type III RNA polymerases, which are processed in the nucleus into relatively small sized (70-100 nucleotides long) precursor miRNAs (pre-miRNA) prior to their export to the cytoplasm where they undergo further processing. Following processing within the cytoplasm, the mature single-stranded miRNAs, in association with / as part of a complex regulatory system (RNA induced silencing complex), play key role in the regulation of gene expression at the post-transcriptional level. Several studies have reported that these interactions with the 'target' may involve direct binding of miRNA to the open reading frames (ORFs)

of the target mRNA sequences or the binding of miRNA to the partial-complementary sequences present at the 5'- or the 3'-untranslated regions (UTRs); later being the most common event¹⁰⁻¹⁴.

MicroRNA as biomarkers of breast cancer

During the last decade, several research groups have reported that miRNA impact the cell proliferation as well as apoptotic mechanisms and thus perform fundamental role in tumorigenesis or cancer progression¹⁵⁻¹⁷. In breast cancer, one of the most prevalent forms of cancer in women worldwide, aberrant expression levels of miRNAs at the different stages of malignancies have been observed. Many different methods were employed for analyzing the expression patterns in these studies which include Northern hybridization (low-throughput technique), microarray, quantitative real-time PCR and next-generation sequencing platforms. By integrating the miRNA expression data with messenger RNA data, new miRNA-gene interactions could be explored which helped in establishing the link between miRNAs

Table 1: Functional microRNAs that are reported to be up- or down-regulated in breast cancer

miRNAs	Status	Functional role in breast cancer	References
hsa-miR-7a	Down-regulated	Suppresses cell proliferation and induces apoptosis	²⁰ Shi et al., 2015
hsa-miR-10b	Upregulated	Metastasis	²² Devulapally et al., 2015
hsa-miR-16	Down-regulated	Apoptosis	²⁴ Ng et al., 2013
Has-miR-20b	Upregulated	Metastasis, cell proliferation	¹⁵ Zhou et al., 2014
hsa-miR-21	Upregulated	Metastasis, cell proliferation	²¹ Si et al., 2013
hsa-miR-23b	Upregulated	Post-transcriptional regulation	²⁵ Fu et al., 2011
hsa-miR-27a	Upregulated	Cell cycle progression, G2-M checkpoint regulation	²⁶ Heneghan et al., 2010
hsa-miR-34a	Upregulated	DNA damage, proliferation	²⁷ Blenkiron et al., 2007
has-miR-125	Down-regulated	Inhibits tumor cell growth and cell invasion	¹⁶ Hsieh et al., 2014
hsa-miR-145	Down-regulated	Suppresses cell invasion and metastasis	²⁷ Blenkiron et al., 2007
hsa-miR-155	Upregulated	TGF-beta signaling	²⁶ Heneghan et al., 2010
hsa-miR-191	Upregulated	Proliferation and migration	²⁸ Ahmad et al., 2013
hsa-miR-195	Upregulated	Apoptosis	²⁶ Heneghan et al., 2010
hsa-miR-200c	Down-regulated	TGF-beta signaling	²⁹ Kayani et al., 2011
hsa-miR-206	Upregulated	ER signaling	³⁰ Kondo et al., 2008
hsa-miR-451	Upregulated	Post-transcription regulation	²⁴ Ng et al., 2013
hsa-miR-491	Down-regulated	Inhibits cell proliferation	²⁰ Shi et al., 2015
hsa-miR-520c	Upregulated	Metastasis, migration, invasion	²⁸ Ahmad et al., 2013
hsa-miR-708	Down-regulated	Inhibits metastasis	¹⁹ Ryu et al., 2013
hsa-miR-888	Upregulated	Metastasis	²³ McCubrey et al., 2014

and breast cancer oncogenesis, invasion and/or metastasis^{17,18}. On the basis of these results, breast-cancer associated miRNAs have been grouped into two main categories: 1) one that activates cell proliferation/ oncogenesis and hence been referred as 'oncomirs' and 2) the others that inhibit/suppress tumor cell progression. Some of these reported miRNAs along with their functional roles in breast cancer have been summarized in Table. 1.

CONCLUSION

All referenced reports have highlighted the potential of miRNAs in differentiating normal and metastatic tumor cells. Major challenge, however, is to validate the candidate miRNAs in large study cohorts. Moreover, due to the complex nature of cancer/neoplasia, a single miRNA is unlikely to serve as predictive biomarker; instead a panel of such tumor- and stage-specific miRNAs should be engaged in breast cancer screening compliance.

REFERENCES

1. Anker, P., Mulcahy, H. and Stroun, M. (2003) Circulating nucleic acids in plasma and serum as a non-invasive investigation for cancer. *Intern. J. Cancer*, 103: 149-152.
2. Brambilla, C., Fievet, F., Jeanmart, M., Fraipont, F.D., Lantvejou, S., Frappat, V., Ferretti, G., Brichan, P.Y. and Sibilot, D.M. (2003) Early detection of lung cancer: role of biomarkers. *Eur Respir*, 21: 36-44.
3. Zabarovsky, E.R., Lerman, M.I. and Minna, J.D. (2002) Tumor suppressor on chromosome 3p involved in the pathogenesis of lung and other cancers. *Oncogene*, 21: 6915-6935.
4. Silva, J.M., Dominguez, G., Garcia, J.M., Gonzalez, R., Villanueva, M.J., Navarro, F., et al. (1999). Presence of tumour DNA in plasma of breast cancer patients: clinicopathological correlations. *Cancer Res*, 59: 3251-3256.
5. Sozzi, G., Conte, D., Leon, M., Cinincione, R., Roz, L., Ratcliffe, C., et al. (2003) Quantification of free circulating DNA as a diagnostic marker in lung cancer. *J Clin Onc*, 21: 3902-3908.
6. Kopreski, M.S., Benko, F.A., Kwak, L.W. and Gocke, C.D. (1999) Detection of tumor messenger RNA in the serum of patient with malignant melanoma. *Clin Cancer Res*, 5: 1961-1965.
7. Li, Y., Elashoff, D., Oh, M., Sinha, U. S., John, M.A.R., Zhou, X., Abemayor, E. and Wong, D.T. (2006) Serum circulating human mRNA profiling and its utility for oral cancer detection. *J Clin Oncol*, 24: 1754-1760.
8. Chen, X.Q., Bannefoi, H., Pelte, M.F., Lyautey, J., Ledderry, C., Movarakhi, S., Schaeffer, P. et al. (2000). Telomerase RNA as a detection marker in the serum of breast cancer patient. *Clin Cancer Res*, 6: 3823-3826.
9. El-Hefnawy, T., Raja, S., Kelly, L., Bigbee, W.L. Kirkwood, J. M., Luketich, J.D. and Godfrey, T. E. (2004) Characterization of amplifiable, circulating RNA in plasma and its potential as a tool for cancer diagnostics. *Clin Chem*, 50: 564-573.
10. Lee, R.C., Feinbaum, R.L., and Ambros, V. (1993). The *C. elegans* heterochronic gene lin-4 encodes small RNAs with antisense complementarity to lin-14. *Cell* 75, 843-854.
11. Etheridge A, Lee I, Hood L, Galas D and Wang K. (2011) Extracellular microRNA: a new source of biomarkers. *Mutational Research*, 717: 85-90.
12. Reinhart, B.J., Slack, F.J., Basson, M., Pasquinelli, A.E., Bettinger, J.C., Rougvie, A.E., Horvitz, H.R., and Ruvkun, G. (2000) The 21-nucleotide let-7 RNA regulates developmental timing in *Caenorhabditis elegans*. *Nature* 403, 901-906.
13. Lee Y, Kim M, Han J, Yeom K.H and Lee S. MicroRNA genes are transcribed by RNA polymerase II. *EMBO J*, 2004; 23: 4051-4060.
14. Borchert G.M, Lanier W and Davidson B.L. (2005) RNA polymerase III transcribes human microRNAs. *Nature of Structural Molecular Biology*, 13: 1097-1101.
15. Zhou, W., Shi, G., Zhang, Q., Wu, Q., Li, B. and Zhang, Z. (2014) MicroRNA-20b promotes cell growth of breast cancer cells partly via targeting phosphatase and tensin homologue (PTEN). *Cell Biosci.*, 4: 62.
16. Hsieh, T., Hsu, C., Tsai, C., Long, C. and Chai C. (2014) miR-125a-5p is a prognostic biomarker that targets HDAC4 to suppress breast tumorigenesis. *Oncotarget*, 6: 494-509.
17. Leivonen, S.K., Sahlberg, K.K., Mäkelä, R., Due, E.U., Kallioniemi, O., Børresen-Dale, A.L. et al. (2014) High-throughput screens identify microRNAs essential for HER2 positive breast cancer cell growth. *Mol. Oncol.*, 8: 93-104.
18. Dhahbi, J.M., Atamna H., Boffelli, D., Magis, W., Spindler, S.R and Martin, D.I. (2011) Deep sequencing reveals novel microRNAs and regulation of microRNA expression during cell senescence. *PLoS ONE*, 6:20509.
19. Ryu, S., McDonnell, K., Choi, H., Gao, D., Hahn, M., Joshi, N. et al. (2013). Suppression of miRNA-708 by polycomb group promotes metastases by calcium-induced cell migration. *Cancer Cell*, 23: 63-76
20. Shi, Y., Luo, X., Li, P., Tan, J., Wang, X., Xiang, X. et al. (2015) miR-7-5p suppresses cell proliferation and induces apoptosis of breast cancer cells mainly by targeting REGY. *Cancer Lett.*, 358: 27-36
21. Si, H., Sun, X., Chen, Y., Cao, Y., Chen, S., Wang, H. et al. (2013) Circulating microRNA-92a and microRNA-21 as minimally invasive biomarkers for primary breast cancer. *J. Cancer Res. Clin. Oncol.*, 139: 223-229.
22. Devulapally, R., Sekar, N.M., Sekar, T.V., Foygel, K., Massoud, T.F., Paulmurugan, R. et al. (2015). Polymer nanoparticles mediated codelivery of anti-miR-10b and anti-miR-21 for achieving triple negative breast cancer novel therapy. *ACS Nano*, 9(3): 2290-2302.
23. McCubrey Ja, Davis, N.M., Abrams, S.L., Montalto, G., Cervello, M., Libra, M. et al. (2014) Targeting breast cancer initiating cells: advances in breast cancer research and therapy. *Adv. Biol. Regul.*, 56: 1-27.

24. Ng, E. K. O., Li, R., Shin, V. Y., Jin, H. C., Leung, C. P. H., Edmond, S. K. M., Pang, R., Chua, D., Chu, K., Law, L. W., Poon, P. T. R., & Kwong, A. (2013). Circulating microRNAs as specific biomarkers for breast cancer detection. *PLoS ONE*, 8(1), e53141.
25. Fu, W. S., Chen, L., & Man, Y. (2011). miRNA biomarkers in breast cancer detection and management. *Journal of Cancer*, 2, 116-122.
26. Heneghan, M. H., Miller, N., Kelly, R., Newell, J., & Kerin, J. M. (2010). Systemic *miRNA-195* differentiates breast cancer from other malignancies and is a potential biomarker for detecting noninvasive and early stage disease. *The Oncologist*, 15, 673–682.
27. Blenkiron, L., Goldstein, D. L., & Thorne, P. N. (2007). MicroRNA expression profiling of human breast cancer identifies new markers of tumor subtype. *Genome Biology*, 8(10), R214.
28. Ahmad, J., Hasnainb, E. S., Siddiquia, A. M., Ahamed, M., Musarrata, J., & Al-Khedhairya, A. A. (2013). MicroRNA in carcinogenesis & cancer diagnostics: A new paradigm. *Indian J Med Res*, 137, 680-694.
29. Kayani, M. R., Kayani, A. M., Malik, A. F., & Faryal, R. (2011). Role of miRNAs in breast cancer. *Asian Pacific J Cancer Prev*, 12, 3175-3180.
30. Kondo, N., Toyama, T., Sugiura, H., Fujii, Y., & Yamashita, H. (2008). *miR-206* expression is down-regulated in estrogen receptor α -positive human breast cancer. *Cancer Research*, 68(13), 5004–5008.